1afo
Evolutionary trace report by report_maker
March 27, 2010

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1 INTRODUCTION

From the original Protein Data Bank entry (PDB id 1afo):

**Title:** Dimeric transmembrane domain of human glycophorin a, nmr, 20 structures

**Compound:** Mol id: 1; molecule: glycophorin a; chain: a, b; fragment: transmembrane peptide; engineered: yes

**Organism, scientific name:** Homo Sapiens;

1afo contains a single unique chain 1afoA (40 residues long) and its homologue 1afoB. This is an NMR-determined structure – in this report the first model in the file was used.

2 CHAIN 1AFOA

2.1 Q03870 overview

From SwissProt, id Q03870, 100% identical to 1afoA:

**Description:** Glycophorin Erik (STA) precursor.

**Organism, scientific name:** Homo sapiens (Human).

**Taxonomy:** Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Catarrhini; Hominidae; Homo.

2.2 Multiple sequence alignment for 1afoA

For the chain 1afoA, the alignment 1afoA.msf (attached) with 25 sequences was used. The alignment was downloaded from the HSSP database, and fragments shorter than 75% of the query as well as duplicate sequences were removed. It can be found in the attachment to this report, under the name of 1afoA.msf. Its statistics, from the alistat program are the following:
2.3 Residue ranking in 1afoA

The 1afoA sequence is shown in Fig. 1, with each residue colored according to its estimated importance. The full listing of residues in 1afoA can be found in the file called 1afoA.ranks_sorted in the attachment.

2.4 Top ranking residues in 1afoA and their position on the structure

In the following we consider residues ranking among top 25% of residues in the protein. Figure 2 shows residues in 1afoA colored by their importance: bright red and yellow indicate more conserved/important residues (see Appendix for the coloring scheme). A Pymol script for producing this figure can be found in the attachment.

2.4.1 Clustering of residues at 25% coverage. Fig. 3 shows the top 25% of all residues, this time colored according to clusters they belong to. The clusters in Fig.3 are composed of the residues listed in Table 1.

<table>
<thead>
<tr>
<th>cluster color</th>
<th>size</th>
<th>member residues</th>
</tr>
</thead>
<tbody>
<tr>
<td>red</td>
<td>9</td>
<td>76, 80, 82, 83, 85, 86, 88, 89, 91</td>
</tr>
</tbody>
</table>

Table 1. Clusters of top ranking residues in 1afoA.

2.4.2 Overlap with known functional surfaces at 25% coverage. The name of the ligand is composed of the source PDB identifier and the heteroatom name used in that file.

Interface with 1afoB. Table 2 lists the top 25% of residues at the interface with 1afoB. The following table (Table 3) suggests possible disruptive replacements for these residues (see Section 3.6).

<table>
<thead>
<tr>
<th>res</th>
<th>type</th>
<th>subst’s</th>
<th>cvg</th>
<th>noc/</th>
<th>dist</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>(%)</td>
<td></td>
<td></td>
<td>Å</td>
</tr>
</tbody>
</table>

Table 2. Residues at the interface with 1afoB. Continued in next column.
Table 2. The top 25% of residues in 1afoA at the interface with 1afoB. (Field names: res: residue number in the PDB entry; type: amino acid type; subst’s: substitutions seen in the alignment; with the percentage of each type in the bracket; noc/bb: number of contacts with the ligand, with the number of contacts realized through backbone atoms given in the bracket; dist: distance of closest approach to the ligand.)

<table>
<thead>
<tr>
<th>res</th>
<th>type</th>
<th>subst’s (%)</th>
<th>cvg</th>
<th>noc/bb</th>
<th>dist (Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>76</td>
<td>I</td>
<td>I(100)</td>
<td>0.07</td>
<td>391/47</td>
<td>1.87</td>
</tr>
<tr>
<td>83</td>
<td>G</td>
<td>G(100)</td>
<td>0.07</td>
<td>124/65</td>
<td>2.35</td>
</tr>
<tr>
<td>82</td>
<td>A(95) L(4)</td>
<td></td>
<td>0.15</td>
<td>30/10</td>
<td>3.31</td>
</tr>
<tr>
<td>91</td>
<td>I(95) L(4)</td>
<td></td>
<td>0.17</td>
<td>76/0</td>
<td>1.98</td>
</tr>
<tr>
<td>86</td>
<td>G(92) L(4)</td>
<td></td>
<td>0.20</td>
<td>2/0</td>
<td>4.97</td>
</tr>
<tr>
<td>80</td>
<td>V(95) P(4)</td>
<td></td>
<td>0.25</td>
<td>277/67</td>
<td>1.80</td>
</tr>
<tr>
<td>88</td>
<td>I(95) M(4)</td>
<td></td>
<td>0.25</td>
<td>29/4</td>
<td>3.18</td>
</tr>
</tbody>
</table>

Table 3. List of disruptive mutations for the top 25% of residues in 1afoA, that are at the interface with 1afoB.

<table>
<thead>
<tr>
<th>res</th>
<th>type</th>
<th>disruptive mutations</th>
</tr>
</thead>
<tbody>
<tr>
<td>76</td>
<td>I</td>
<td>(YR) (TH) (SKECG) (FQWD)</td>
</tr>
<tr>
<td>83</td>
<td>G</td>
<td>(KER) (FQMWHD) (NYLPI) (SVA)</td>
</tr>
<tr>
<td>82</td>
<td>A</td>
<td>(YR) (KE) (H) (QD)</td>
</tr>
<tr>
<td>91</td>
<td>I</td>
<td>(YR) (TH) (SKECG) (FQWD)</td>
</tr>
<tr>
<td>86</td>
<td>G</td>
<td>(R) (KE) (H) (Y)</td>
</tr>
<tr>
<td>80</td>
<td>V</td>
<td>(YR) (KE) (H) (QD)</td>
</tr>
<tr>
<td>88</td>
<td>I</td>
<td>(Y) (R) (TH) (SCG)</td>
</tr>
</tbody>
</table>

Figure 4 shows residues in 1afoA colored by their importance, at the interface with 1afoB.

2.4.3 Possible novel functional surfaces at 25% coverage. One group of residues is conserved on the 1afoA surface, away from (or substantially larger than) other functional sites and interfaces recognizable in PDB entry 1afo. It is shown in Fig. 5. The residues belonging to this surface "patch" are listed in Table 4, while Table 5 suggests possible disruptive replacements for these residues (see Section 3.6).

Table 4. Residues forming surface "patch" in 1afoA.

<table>
<thead>
<tr>
<th>res</th>
<th>type</th>
<th>substitutions (%)</th>
<th>cvg</th>
</tr>
</thead>
<tbody>
<tr>
<td>76</td>
<td>I</td>
<td>I(100)</td>
<td>0.07</td>
</tr>
<tr>
<td>83</td>
<td>G</td>
<td>G(100)</td>
<td>0.07</td>
</tr>
<tr>
<td>85</td>
<td>I(95) L(4)</td>
<td></td>
<td>0.12</td>
</tr>
<tr>
<td>89</td>
<td>L(95) P(4)</td>
<td></td>
<td>0.12</td>
</tr>
<tr>
<td>82</td>
<td>A(95) L(4)</td>
<td></td>
<td>0.15</td>
</tr>
</tbody>
</table>

Continued in next column

Table 4. continued

<table>
<thead>
<tr>
<th>res</th>
<th>type</th>
<th>substitutions (%)</th>
<th>cvg</th>
</tr>
</thead>
<tbody>
<tr>
<td>91</td>
<td>I(95) L(4)</td>
<td></td>
<td>0.17</td>
</tr>
<tr>
<td>86</td>
<td>G(92) L(4)</td>
<td></td>
<td>0.20</td>
</tr>
<tr>
<td>80</td>
<td>V(95) P(4)</td>
<td></td>
<td>0.25</td>
</tr>
<tr>
<td>88</td>
<td>I(95) M(4)</td>
<td></td>
<td>0.25</td>
</tr>
</tbody>
</table>

Fig. 4. Residues in 1afoA, at the interface with 1afoB, colored by their relative importance. 1afoB is shown in backbone representation (See Appendix for the coloring scheme for the protein chain 1afoA.)

Fig. 5. A possible active surface on the chain 1afoA.
3 NOTES ON USING TRACE RESULTS

3.1 Coverage
Trace results are commonly expressed in terms of coverage: the residue is important if its “coverage” is small - that is if it belongs to some small top percentage of residues [100% is all of the residues in a chain], according to trace. The ET results are presented in the form of a table, usually limited to top 25% percent of residues (or to some nearby percentage), sorted by the strength of the presumed evolutionary pressure. (I.e., the smaller the coverage, the stronger the pressure on the residue.) Starting from the top of that list, mutating a couple of residues should affect the protein somehow, with the exact effects to be determined experimentally.

3.2 Known substitutions
One of the table columns is “substitutions” - other amino acid types seen at the same position in the alignment. These amino acid types may be interchangeable at that position in the protein, so if one wants to affect the protein by a point mutation, they should be avoided. For example if the substitutions are “RVK” and the original protein has an R at that position, it is advisable to try anything, but RVK. Conversely, when looking for substitutions which will not affect the protein, one may try replacing, R with K, or (perhaps more surprisingly), with V. The percentage of times the substitution appears in the alignment gives the number of contacts heavy atoms of the residue in question make across the interface, as well as how many of them are realized through the backbone atoms (if all or most contacts are through the backbone, mutation presumably won’t have strong impact). Two heavy atoms are considered to be “in contact” if their centers are closer than 5Å.

3.3 Surface
To detect candidates for novel functional interfaces, first we look for residues that are solvent accessible (according to DSSP program) by at least 10Å², which is roughly the area needed for one water molecule to come in the contact with the residue. Furthermore, we require that these residues form a “cluster” of residues which have neighbor within 5Å from any of their heavy atoms.

Note, however, that, if our picture of protein evolution is correct, the neighboring residues which are not surface accessible might be equally important in maintaining the interaction specificity - they should not be automatically dropped from consideration when choosing the set for mutagenesis. (Especially if they form a cluster with the surface residues.)

3.4 Number of contacts
Another column worth noting is denoted “noc/bb”; it tells the number of contacts heavy atoms of the residue in question make across the interface, as well as how many of them are realized through the backbone atoms (if all or most contacts are through the backbone, mutation presumably won’t have strong impact). Two heavy atoms are considered to be “in contact” if their centers are closer than 5Å.

3.5 Annotation
If the residue annotation is available (either from the pdb file or from other sources), another column, with the header “annotation” appears. Annotations carried over from PDB are the following: site (indicating existence of related site record in PDB ), S-S (disulfide bond forming residue), hb (hydrogen bond forming residue), jb (james bond forming residue), and sb (for salt bridge forming residue).

3.6 Mutation suggestions
Mutation suggestions are completely heuristic and based on complementarity with the substitutions found in the alignment. Note that they are meant to be disruptive to the interaction of the protein with its ligand. The attempt is made to complement the following properties: small [AVGSTC], medium [LPNQDEM1K], large [W FYH R], hydrophobic [LPV AM W F I], polar [G T C Y]; positively [K HR], or negatively [D E] charged, aromatic [W FY H], long aliphatic chain [ERQKM], OH-group possession [SDETY], and NH2 group possession [NQRK]. The suggestions are listed according to how different they appear to be from the original amino acid, and they are grouped in round brackets if they appear equally disruptive. From left to right, each bracketed group of amino acid types resembles more strongly the original (i.e. is, presumably, less disruptive) These suggestions are tentative - they might prove disruptive to the fold rather than to the interaction. Many researcher will choose, however, the straightforward alanine mutations, especially in the beginning stages of their investigation.

4 APPENDIX

4.1 File formats
Files with extension “ranks_sorted” are the actual trace results. The fields in the table in this file:

- alignment# number of the position in the alignment
- residue# residue number in the PDB file
- type amino acid type
- rank rank of the position according to older version of ET
- variability has two subfields:
  1. number of different amino acids appearing in in this column of the alignment
  2. their type
- rho ET score - the smaller this value, the lesser variability of this position across the branches of the tree (and, presumably, the greater the importance for the protein)
- cvg coverage - percentage of the residues on the structure which have this rho or smaller
- gaps percentage of gaps in this column
4.2 Color schemes used

The following color scheme is used in figures with residues colored by cluster size: black is a single-residue cluster; clusters composed of more than one residue colored according to this hierarchy (ordered by descending size): red, blue, yellow, green, purple, azure, turquoise, brown, coral, magenta, LightSalmon, SkyBlue, violet, gold, bisque, LightSlateBlue, orchid, RosyBrown, MediumAquamarine, DarkOliveGreen, CornflowerBlue, grey55, burlywood, LimeGreen, tan, DarkOrange, DeepPink, maroon, BlanchedAlmond.

The colors used to distinguish the residues by the estimated evolutionary pressure they experience can be seen in Fig. 6.

4.3 Credits

4.3.1 Alistat

alistat reads a multiple sequence alignment from the file and shows a number of simple statistics about it. These statistics include the format, the number of sequences, the total number of residues, the average and range of the sequence lengths, and the alignment length (e.g. including gap characters). Also shown are some percent identities. A percent pairwise alignment identity is defined as (idents / MIN(len1, len2)) where idents is the number of exact identities and len1, len2 are the unaligned lengths of the two sequences. The "average percent identity", "most related pair", and "most unrelated pair" of the alignment are the average, maximum, and minimum of all (N)(N-1)/2 pairs, respectively. The "most distant seq" is calculated by finding the maximum pairwise identity (best relative) for all N sequences, then finding the minimum of these N numbers (hence, the most outlying sequence). alistat is copyrighted by HHMI/Washington University School of Medicine, 1992-2001, Sander and MPI-MF, 1983, 1985, 1988, 1994 1995, CMBI version by Elmar.Krieger@.cmbi.kun.nl November 18,2002, http://www.cmbi.kun.nl/gv/dssp/descrip.html.

4.3.2 CE

To map ligand binding sites from different source structures, report_maker uses the CE program: http://cl.sdsc.edu/. Shindyavov IN, Bourne PE (1998) "Protein structure alignment by incremental combinatorial extension (CE) of the optimal path". Protein Engineering 11(9) 739-747.

4.3.3 DSSP

In this work a residue is considered solvent accessible if the DSSP program finds it exposed to water by at least 10Å², which is roughly the area needed for one water molecule to come in contact with the residue. DSSP is copyrighted by W. Kabsch, C. Sander and MPI-MF, 1983, 1985, 1988, 1994 1995, CMBI version by Elmar.Krieger@cmbi.kun.nl November 18,2002, http://www.cmbi.kun.nl/gv/dssp/descrip.html.

4.3.4 HSSP


4.3.5 LaTeX

The text for this report was processed using L\textup{\La}T\textup{E}X; Leslie Lamport, “LaTeX: A Document Preparation System Addison-Wesley,” Reading, Mass. (1986).

4.3.6 Muscle


4.3.7 Pymol

The figures in this report were produced using PyMol. The scripts can be found in the attachment. PyMol is an open-source application copyrighted by DeLano Scientific LLC (2005). For more information about PyMol see http://pymol.sourceforge.net/. (Note for Windows users: the attached package needs to be unzipped for PyMol to read the scripts and launch the viewer.)

4.4 Note about ET Viewer

Dan Morgan from the Lichtarge lab has developed a visualization tool specifically for viewing trace results. If you are interested, please visit:

http://mammoth.bcm.tmc.edu/traceview/

The viewer is self-unpacking and self-installing. Input files to be used with ETV (extension .etvx) can be found in the attachment to the main report.

4.5 Citing this work


4.6 About report_maker

report_maker was written in 2006 by Ivana Mihalek. The 1D ranking visualization program was written by Ivica Reš. report_maker is copyrighted by Lichtarge Lab, Baylor College of Medicine, Houston.
4.7 Attachments

The following files should accompany this report:

- 1afoA.complex.pdb - coordinates of 1afoA with all of its interacting partners
- 1afoA.etvx - ET viewer input file for 1afoA
- 1afoA.cluster_report.summary - Cluster report summary for 1afoA
- 1afoA.ranks - Ranks file in sequence order for 1afoA
- 1afoA.clusters - Cluster descriptions for 1afoA
- 1afoA.msf - the multiple sequence alignment used for the chain 1afoA
- 1afoA.descr - description of sequences used in 1afoA msf
- 1afoA.ranks_sorted - full listing of residues and their ranking for 1afoA
- 1afoA.1afoB.if.pml - Pymol script for Figure 4
- 1afoA.cbcvg - used by other 1afoA – related pymol scripts